

SYNTHESIS AND BIOLOGICAL EVALUATION OF CRYPTOPHYCIN ANALOGS WITH SUBSTITUTION AT C-6 (FRAGMENT C REGION)

David L. Varie,* Chuan Shih, David A. Hay, Sherri L. Andis, Tom H. Corbett, Lynn S. Gossett, Samantha K. Janisse, Michael J. Martinelli, Eric D. Moher, Richard M. Schultz, and John E. Toth

Lilly Research Laboratories, A Division of Eli Lilly and Co., Indianapolis, IN 46285, U.S.A. and

*Karmanos Cancer Institute, Wayne State University, Detroit, MI 48202, U.S.A.

Abstract: Analogs of the antitumor agents cryptophycins 1 and 8 with dialkyl substitution at C-6 (fragment C) were synthesized and evaluated for in vitro cytotoxicity against human leukemia cells (CCRF-CEM). The activity of these analogs decreased as the size of the substituents at C-6 increased. The C-6 spirocylopropyl compound (2g) was highly potent in vitro and showed excellent antitumor activity in animal models. © 1999 Elsevier Science Ltd. All rights reserved.

Received 2 November 1998; accepted 14 December 1998

The depsipeptide cryptophycins derived from terrestrial blue-green algae exhibit high activity against a broad spectrum of solid tumors. The major cytotoxic component of the algal extracts, cryptophycin 1 (1a), was first isolated from a *Nostoc* cyanobacterium by Schwartz and coworkers in 1990. Moore and coworkers later isolated several cryptophycins, including 1a, from *Nostoc* sp GSV 224. Following the report of the total synthesis of cryptophycin 1 by Tius et al., several syntheses of cryptophycins were reported. A number of synthetic cryptophycins with promising antitumor activity, including the C-6 dimethyl substituted cryptophycins 52 (1b) and 55 (2b), have subsequently been prepared.

Previous reports have shown that acyclic analogs of the cryptophycins (e.g., cryptophycin 5, 3) in which the C-5 ester bond has been cleaved, are 3–4 orders of magnitude less active against human tumor cell lines in vitro.⁶ We hypothesized that hydrolysis of this ester bond may be one process that would decrease the cytotoxicity of cryptophycins. If so, increased steric bulk at C-6 (geminal disubstitution, for example) may result in compounds with improved stability and antitumor activity. This report describes the preparation and

biological evaluation of eleven novel cryptophycin epoxide (1) and chlorohydrin (2) analogs having dialkyl substituents at C-6.

Retrosynthetic analysis of the cryptophycins provides four fragments (A–D), which can be independently synthesized and coupled to form the macrocycle. Fragments A–B (12) and D (9) were prepared as previously described by Tius.³ The new fragment C–D derivatives (10) were prepared as shown in Schemes 1 and 2. Ethyl cyanoacetate (4) was treated with Cs_2CO_3 and the appropriate alkyl halide in DMF to provide the dialkyl nitriles 5c-f. The ester and nitrile groups were reduced with LiAlH₄ to give amino alcohols 6c-f. Protection of the primary amine as the t-butyloxycarbonyl (Boc) derivative and oxidation of the primary alcohol with catalytic ruthenium tetroxide⁷ yielded the desired fragment C acids 8c-f.

Scheme 1. Preparation of Fragment C-D Analogs 10c-f.

Reagents. (a) Cs₂CO₃/DMF/alkyl halide (Etl, Prl, Br(CH₂)₃Br, Br(CH₂)₄Br)/0-25 $^{\circ}$ C. (b) LiAlH₄/THF. (c) Boc₂O/NaOH/THF. (d) cat. RuCl₃/NalO₄/CH₃CN/CCl₄/H₂O. (e) i. CDI; ii. 9/THF or PhCH₃/reflux. (f) 0.2 mol% Pd (Ph₃P)₄/morpholine/THF.

Acids 8 were reacted with 1,1-carbonyldiimidazole (CDI) and treated with the allyl ester of (S)-leucic acid (Fragment D, 9) in refluxing THF or toluene to give the fragment C-D esters.⁸ The allyl protecting group was

removed with catalytic Pd(PPh₃)₄ in the presence of morpholine.⁹ The cyclopropyl substituted fragment C–D analog 10g was prepared from commercially available 2-cyclopropyl cyanoacetic acid (11, Scheme 2).

Scheme 2. Preparation Cyclopropyl Fragment C-D Acid 10g.

Reagents. (a) $H_2/Pt_2O/HOAc$ (86%). (b) $Boc_2O/NaOH/dioxane$ (93%). (c) $DCC/9/CH_2Cl_2$ (70%). (d) cat. $Pd(PPh_3)_4/morpholine/THF$ (95%).

Scheme 3. Preparation of Cryptophycin Analogs 1 and 2.

$$\begin{array}{c} \text{Me} \\ \text{Me} \\ \text{O} \\$$

Reagents. (a) DCC/DMAP/CH $_2$ Cl $_2$. (b) TFA. (c) 1 equiv 2-hydroxypyridine/PhCH $_3$. d) m-CPBA/CHCl $_3$ /rt. (e) i. TMSCl/CHCl $_3$ /-60 $^{\circ}$ C; ii. MeOH; iii. chromatography. (f) K $_2$ CO $_3$ /MeCN/H $_2$ O.

Fragment C–D acids 10 were then coupled to fragment A–B (12) using 1,3-dicyclohexylcarbodiimide (DCC) and DMAP (Scheme 3). Removal of the Boc group with TFA followed by macrolactamization promoted by 2-hydroxypyridine¹⁰ gave the cryptophycin macrocycles 14c–g. The styrene double bond was epoxidized with m-CPBA to provide a 2:1 β : α mixture of diastereomeric epoxides 15, which were not separated. The epoxide mixtures were converted to the chlorohydrins with trimethylsilyl chloride (TMSCI) at -60 °C and the desired chlorohydrins 2c–g were isolated by silica gel or reverse phase chromatography in 10–31% overall yield from 14.¹¹ Pure β -epoxides 1c–g were obtained from the chlorohydrins by reaction with K,CO₃ in acetonitrile/water.

Compounds 1 and 2 were tested for in vitro cytotoxicity against the CCRF-CEM human leukemia cell line.¹² As can be seen in Table 1, as the size of the alkyl substituents at C-6 increases, the IC₅₀ value markedly decreases for both the epoxide and chlorohydrin compounds. In the epoxide series, the spirocyclopropyl compound (1g), which is tenfold less active than cryptophycin 52 (1b), is the most active of the new analogs. In the chlorohydrin series, the cyclopropyl analog 2g is approximately three times more active than cryptophycin 55 (2b); all other analogs are at least tenfold less active than cryptophycin 55.

Table 1. Cytotoxicity of Cryptophycin Fragment C Analogs 1 and 2 (CCRF-CEM Cell L	Table 1.	Cytotoxicity of	Cryptophycin Fragment	C Analogs 1 and 2	(CCRF-CEM Cell Line
---	----------	-----------------	-----------------------	-------------------	---------------------

Compound	R	IC ₅₀ (nM)	IC ₅₀ (nM)
		1 (epoxide)	2 (chlorohydrin)
b	Me	0.02	0.05
c	Et	1.1	1.6
d	Pr	8.5	8.7
e	$(CH_2)_4$	0.3	0.4
f	(CH ₂) ₅	63	110
g	$(CH_2)_2$	0.16	0.014

Seeking to improve the hydrolytic stability further, the spirocyclopropyl amide derivative (20) was prepared using N-Boc L-leucine (16) as the fragment D building block (Scheme 4). Thus fragment A-B (12) was coupled with 16 using DCC to provide ester 17 in 80% yield. The Boc group of 17 was removed with TFA, and the resulting amine was reacted with acid 8g in the presence of 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide (EDCI) and 1-hydroxybenzotriazole (HOBT) to give compound 18 in 77% yield. The macrolactam 19 was formed by sequential removal of the trichloroethyl ester (Zn/HOAc) and Boc protecting groups of 18 followed by ring closure using pentafluorophenyl diphenylphosphinate (FDPP) as described by Tius.³ Epoxidation of 19 with m-CPBA gave a 2:1 mixture of β : α epoxides which were converted directly to the chlorohydrins with TMSCl. The desired chlorohydrin diastereomer (20) was isolated by chromatography on silica gel.

Scheme 4. Preparation of Cyclopropyl Amide 20.

Reagents. (a) **16**, DCC/DMAP/CH₂Cl₂ (80%). (b) TFA. (c) EDCI/HOBt/**8g** (77%). (d) Zn/HOAc. (e) TFA (81%). (f) Et₂Ni-Pr/FDPP/DMF (82%). (g) m-CPBA/CH₂Cl₂ (90%). (h) i. TMSCI/CHCl₃/-60 $^{\circ}$ C; ii. chromatography (35%).

Compound 20 was tested for cytotoxicity against the CCRF-CEM cell line. The results (Table 2) indicate that replacing the fragment C-D ester with an amide has minimal effect on the cytotoxicity. The IC₅₀ value for amide 20 is nearly identical to that for the ester 2g and the previously reported 6,6-dimethyl amide 21.¹³

Compounds 2d, 2g, and 20 were evaluated for in vivo antitumor activity (murine pancreatic adenocarcinoma PO3 model).⁶ As might be expected, the C-6 dipropyl analog (2d) was essentially inactive at all doses tested (T/C >90% vs. T/C <5% for 2b). This result is consistent with the cell-based cytotoxicity assay results. However, the C-6 cyclopropyl compounds 2g and 20, which were highly potent in vitro, showed excellent anti-tumor activity in vivo (T/C of 3-7% and 36%, respectively). It is interesting to note that the presumably more stable amide 20 is slightly less active than the corresponding ester analog 2g.

These studies show that shielding substituents larger than methyl or cyclopropyl at C-6 markedly decrease the biological activity of cryptophycin analogs. While stability of the C-5 ester bond in cryptophycins may be essential for useful antitumor activity, small changes in the steric (and possibly

conformational) features of the fragment C region of the molecule significantly affect the biological activity of these compounds as well.

Table 2. Comparison of Cytotoxicity of Ester and Amide Cryptophycin Analogs.

Compound	X	R	IC ₅₀ (nM)
2b	0	Me	0.05
2g	O	$(CH_2)_2$	0.014
20	NH	$(CH_2)_2$	0.010
21 ¹³	NH	Me	0.016

References and Notes

- 1. Schwartz, R. E.; Hirsch, C. F.; Sesin, D. F.; Flor, J. E.; Chartrain, M.; Fromtling, R. E.; Harris, G. H.; Salvatore, M. J.; Liesch, J. M.; Yudin, K. J. Ind. Microbiol. 1990, 5, 113.
- (a) Subbaraju, G.V.; Golakoti, T.; Patterson, G. M.; Moore, R. E. J. Nat. Prod. 1997, 60, 302. (b)
 Trimurtulu, G.; Ohtani, I.; Patterson, G. M.; Moore, R. E.; Corbett, T. H.; Valeriote, F. A.; Demchik, L. J.
 Am. Chem. Soc. 1994, 116, 4729.
- 3. Barrow, R. A.; Hemscheidt, T.; Liang, J.; Paik, S.; Moore, R. E.; Tius, M. A. J. Am. Chem. Soc. 1995, 117, 2479.
- (a) de Muys, J-M.; Erg, R.; Nguyen, D.; Go, B.; Fortin, S.; Lavallee, J-F. Bioorg. Med. Chem. Lett. 1996, 6, 1111.
 (b) Rej, R.; Nguygen, D.; Go, B.; Fortin, S.; Lavallee, J-F. J. Org. Chem. 1996, 61, 6289.
 (c) Salamonczyk, G. M.; Han, K.; Guo, Z-W.; Sih, C. J. J. Org. Chem. 1996, 61, 6893.
 (d) Ali, S. M.; Goerg, G. I. Tetrahedron Lett. 1997, 38, 1703.
 (e) Leahy, J. W.; Gardinier, K. M. J. Org. Chem. 1997, 62, 7098.
 (f) Shimizu, I.; Furuyama, M. Tetrahedron: Asymmetry 1998, 9, 1351.
- (a) Worzolla, J. F.; Cao, J.; Ehlhardt, W. J.; Harrison, S. D.; Law, K. L.; Martinelli, M. J.; Self, T. D.; Starling, J. J.; Shih, C.; Theobold, K. S.; Toth, J. E.; Zimmerman, J. L.; Corbett, T. H. Proc. Am. Assoc. Cancer Res. 1997, 38:A1516, 225. (b) Corbett, T.; Valeriote, F.; Simpson, C.; Moore, R.; Tius, M.; Hemscheidt, T.; Liang, J.; Paik, S.; Polin, L.; Pugh, S.; Kushner, J.; Harrison, S.; Shih, C.; Martinelli, M. Proc. Am. Assoc. Cancer Res. 1997, 38:A1515, 225.
- Golakoti, T.; Ogino, J.; Heltzel, C. E.; Husebo, T. Le; Jensen, C. M.; Larsen, L. K.; Patterson, G. M. L.; Moore, R. E.; Mooberry, S. L.; Corbet, T. H.; Valeriote, F. A. J. Am. Chem. Soc. 1995, 117, 12030.
- 7. Carlsen, P. H.; Katsuki, T.; Martin, V. S.; Sharpless, K. B. J. Org. Chem. 1981, 46, 3936.
- 3. The reaction of the imidazolide derived from acids 10 and fragment D (9) required higher temperatures and longer reaction times as the size of the alkyl groups α to the C-5 carbonyl increased. Toluene was used as solvent for the preparation of 10d.
- 9. Jeffrey, P. D.; McCombie, S. W. J. Org. Chem. 1982, 47, 587.
- 10. The major by-product of the cyclization reaction was a cyclic dimer. For optimized reaction conditions see: Fray, A. H. *Tetrahedron: Asymmetry* 1998, 9, 2777.
- 11. The diastereomeric mixtures of epoxides 15 were isolated in 80–90% yields. The overall yields of chlorohydrins 2 are based on the amount of pure desired diastereomer obtained after chromatography.
- 12. Schultz, R. M.; Shih C.; Wood, P. G.; Harrison, S. D.; Elhhardt, W. J. Oncology Reports 1998, 5, 1089.
- 13. Norman, B. H.; Hemscheidt, T.; Schultz, R. M.; Andis, S. L. J. Org. Chem. 1998, 63, 5288.